Sampling for Plant-Parasitic Nematodes
Identification and Diagnosis
Department of Plant Pathology, University of Georgia
(Revised 1/2020)

Contact Information
Extension Nematology Laboratory
Department of Plant Pathology
University of Georgia
2350 College Station Road
Athens, GA 30602
Phone: (706) 542-9144
Fax: (706) 542-5957

Dr. Ganpati B. Jagdale (gbjagdal@uga.edu)
Katherine Martin (katherine.martin28@uga.edu)

Charges and Fees (as of 1/1/2020)
$15.00 = All nematode samples submitted through UGA Extension county office. Please provide name of the County of origin and name of Agent as indicated on the form. Results will be submitted to the respective County Office and e-mailed to each client.

$15.00 = Samples from UGA research or demonstration projects. Charge for UGA research samples will be handled by internal billing to UGA chart-strings.

$15.00 = State Certification (Georgia Department of Agriculture).

$30.00 = All in-state nematode samples not submitted via UGA Extension county office.

$60.00 = All nematode samples submitted from out of state.

$100.00 = Identification of free-living nematodes at genus level (call to arrange payment at 706-542-9144).

Payment (or UGA chart-string to be billed) must accompany all samples. Alternatively, sample processing fees can be paid by credit card using the following link: https://estore.uga.edu/C27063_ustores/web/classic/store_main.jsp?STOREID=269&SINGLESTORE=true
A QUICK GUIDE TO SAMPLING FOR NEMATODES

By Ganpati Jagdale

WHAT ARE NEMATODES?
- Nematodes are thread-like microscopic worms that live in all habitats especially soil and water. Most species are beneficial but some are harmful parasites of plants and animals.

WHAT ARE PLANT-PARASITIC NEMATODES?
- Plant-parasitic nematodes are characterized by having a needle-like structure in the mouth called a stylet that depending on the nematode species is used to feed on plant parts that may include roots, stems, leaves and seeds.

WHY ARE PLANT-PARASITIC NEMATODES IMPORTANT?
- Plant-parasitic nematodes can cause significant economic damage to almost all vegetable and field crops, fruit trees, ornamentals and turfgrass by reducing both yield and quality.
- Annual global yield loss caused by nematodes is $78-80 billion with an estimated annual losses in the U.S. of $8 billion.
- Damage caused by nematodes may provide an infection site for other disease-causing pathogens that may cause additional loss in crop quality and yield.

WHAT DOES DAMAGE BY PLANT-PARASITIC NEMATODES LOOK LIKE?
- Damage caused by plant-parasitic nematodes can initially appear as nutrient deficiency in the foliage and therefore is often incorrectly diagnosed.
- As root damage is common due to feeding by plant-parasitic nematodes, nutrients and water essential for proper plant growth may not be absorbed.

HOW ARE PLANT-PARASITIC NEMATODES MANAGED?
- Detection and correct identification of nematode species is the first step in managing plant-parasitic nematodes to check their spread and prevent economic damage.
- As most plant-parasitic nematodes live in the soil associated with plant roots, timely sampling of soil and plant samples is an important step in implementing appropriate strategies for control and preventing economic damage.
- The specific strategy for managing plant-parasitic nematodes varies with the crop.
**HOW TO SAMPLE**

- **Timing:**
  
  The timing of nematode sampling is important because nematode populations fluctuate throughout the year. Nematodes may be undetectable during the winter and early spring but often increase to a very high population density in the early fall before crop harvest when living roots are present. After harvest, nematode population may decline sharply. Thus, taking samples at the appropriate time for a crop decreases the risk of failing to detect a damaging nematode species and helps avoid or reduce the potential of nematode problems in the future. Sampling in the early fall facilitates decision making in implementing appropriate nematode management practices before the next crop is planted.

**The optimum time to take samples for nematode assay from various Georgia crops is given below:**

<table>
<thead>
<tr>
<th>Crop</th>
<th>When to sample</th>
<th>Sampling depth in inches (cm)</th>
<th>Common nematodes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cotton</td>
<td>Oct. and Nov.</td>
<td>8 inches (20)</td>
<td>Lance, Reniform, Root-knot</td>
</tr>
<tr>
<td>Peaches and other fruit orchards</td>
<td>Feb. to Apr. and Sept. to Oct.</td>
<td>8 inches (20)</td>
<td>Root-knot</td>
</tr>
<tr>
<td>Peanuts</td>
<td>Sept. to Oct.</td>
<td>8 inches (20)</td>
<td>Root-knot</td>
</tr>
<tr>
<td>Soybeans</td>
<td>Sept. to Oct.</td>
<td>8 inches (20)</td>
<td>Lance, Reniform, Root-knot, Cyst</td>
</tr>
<tr>
<td>Tobacco</td>
<td>July</td>
<td>8 inches (20)</td>
<td>Root-knot</td>
</tr>
<tr>
<td>Warm season turfgrass</td>
<td>Jun. to Aug.</td>
<td>6 inches (15)</td>
<td>Lance, Reniform, Root-knot, Sting</td>
</tr>
<tr>
<td>Cool season turfgrass</td>
<td>Sept. to Oct.</td>
<td>6 inches (15)</td>
<td>Lance, Reniform, Root-knot, Sting</td>
</tr>
<tr>
<td>Vegetables</td>
<td>Aug. to Sept.</td>
<td>8 inches (20)</td>
<td>Root-knot</td>
</tr>
</tbody>
</table>

**WHAT AND WHERE TO SAMPLE**

In the field, nematode distribution is always patchy because their population density is affected by the crop grown, presence of weed hosts, chemicals used and environmental factors. Because of the patchy distribution of nematodes, it is very important that the soil sample be truly representative of the area sampled. The only way to ensure the sample is representative is to collect soil from many spots around the field rather than from only one or two spots. Soil samples should only be collected when soil moisture is suitable for working the field. Avoid sampling under excessively dry, wet or frozen soil conditions. Soil samples should be taken from around the plant root (rhizosphere). Sampling depth may differ for different crops (see above table). In a fallow field, samples should be taken from the depth of the root zone of the future crop. For fields left fallow, or fields under dry or frozen conditions for a long period of time, the depth of sampling should be more than 20 inches because nematodes tend move deep in the soil to avoid unfavorable conditions.

In cases where there may be infection of the plant by a nematode, samples should be collected from plant parts (leaves, stems, bulbs, tubers or roots) that are suspected or showing symptoms of nematode infection. Both soil and plant samples should be taken from areas showing nematode damage (symptoms) and from healthy areas for comparison.
If a problem area is being sampled, collect soil from the margin of the affected area to ensure presence of high nematode populations (Fig. 1). Even if a small problem area is sampled, soil should still be collected from multiple spots within the area sampled. If a very large area (> 5 acres) is sampled, high-population density areas will be diluted by low population density areas so that areas with nematode problems will be more difficult to identify. Therefore, one composite soil sample should be taken for every 4 to 5 acres. The sample may represent a section that has homogeneous soil type and conditions and is uniformly farmed. The shape of a field may influence the number of acres that a sample represents.

HOW TO SAMPLE

- Properly taken samples from small field units can reduce production costs by allowing the grower to eliminate nematode control practices where they are not needed and implement control practices where they are needed.
- Improper sampling can lead to poor recommendations and economic losses which could have been avoided.
- Both large areas (fields, golf course fairways and rough areas) and small areas (home lawns, recreational parks and golf course greens) should be sampled in a systematic zigzag pattern (Fig. 2A and B).

- As nematodes may not survive in the upper 1-2 inches of soil due to extreme environmental conditions (hot and cold), first remove the upper soil layer and then take samples using a 2.5-cm (1-inch) diameter soil probe (Fig. 3).
- Take 20-30 soil cores in a zigzag pattern from regularly spaced locations throughout a large area (4-5 acre section) or a small area (≤ 1.25 acres [≤5000 m²]).
- Single plants such as fruit trees and ornamental shrubs should be sampled under the canopy drip line (Fig. 4).
- The number of soil cores taken will depend on the size of tree/shrub canopy. For example, about 8 soil cores should be collected for a 10-ft diameter canopy and about 12 soil cores for a 15-ft canopy.
- Collected soil cores should be thoroughly mixed into one composite sample.
- From each thoroughly mixed composite sample, about 1 pint (1/2 liter) of soil should be transferred into a labeled plastic bag for nematode assay.

HANDLING OF SOIL SAMPLES FOR NEMATODE ASSAY
- Improper handling of samples can lead to poor estimation of the nematode population, which in turn could lead to a wrong recommendation.
- Nematodes must be alive for the extraction procedure. Do not allow samples to sit in direct sunlight in the field or in a hot vehicle for even a short time. Excessive heat or drying can kill the nematodes, which will result in failure to detect nematodes in the sample.
- During transit, samples should be stored in an insulated cooler to avoid getting them hot or dry.
- Once in the laboratory, samples can be stored in a refrigerator until they are processed for nematode extraction.

LABELING OF SOIL SAMPLES FOR NEMATODE ASSAY
- Always label plastic bags on the outside with a permanent marker. The label should include sample name/number, location and date of sampling.
- Each sample should be accompanied by a completed NEMATODE ASSAY FORM, which can be downloaded from https://plantpath.caes.uga.edu/content/dam/caes-subsite/plant-pathology/extension-pdfs/Nematode%20Assay%20Form%202020.pdf
- Provide all information requested. The information on this form will determine the usefulness of the resulting recommendations.
- List present, past, and future crop to assist in identifying nematode problems and making management recommendations.
- Also list crop variety grown. Variety (cultivar) information is critical for many crops.
SHIPPING OF SOIL SAMPLES FOR NEMATODE ASSAY

- Deliver in person or mail sample using any express delivery service (FedEx or UPS, U.S. Postal Service etc.).
- Send samples early in the week so that they do not spend the weekend in transit.
- Samples for nematode assay should be submitted through your local county Extension office.
- Your county Extension office will send the samples to the Extension Nematology Laboratory.
- The sender is responsible for all shipping expenses.

Mailing Address:
Extension Nematology Laboratory
Department of Plant Pathology
2350 College Station Road
Athens, GA 30602

WHEN YOU OBTAIN THE RESULTS

- The results of the assay and management recommendations will be returned to you through your county Extension office within 4-5 days after receipt of samples.
- Keep a record of which nematodes are found in which fields.

Nematode disease diagnosis
Ganpati B. Jagdale
Extension Nematology Laboratory
Department of Plant Pathology
University of Georgia
2350 College Station Rd.
Athens, GA 30602
Phone: 706-542-9144
Fax: 706-542-5957
gbjagdal@uga.edu

Administrative and related issues
Dr. Harald Scherm
Professor & Department Head
Department of Plant Pathology
University of Georgia
Athens, GA 30602-7274
Phone: 706-542-2571
Fax: 706-542-1262
scherm@uga.edu